

Course Specifications

Course Title:	Clinical Immunology
Course Code:	373311-2
Program:	Bachelor in Clinical Laboratory Sciences; Level-6 (Program Code: 373000)
Department:	Clinical Laboratory Sciences Department
College:	College of Applied Medical Sciences
Institution:	Taif University







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A. Course Identification

1.	1. Credit hours: 2 Hours Theory			
2.	Course type			
a.				
b.	Required Elective			
3.	3. Level/year at which this course is offered: 7 th Level/ Third Year			
4.	4. Pre-requisites for this course (if any): Basic of Immunology/ 373239-3			
5.	5. Co-requisites for this course (if any): None			

6. Mode of Instruction (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1	Traditional classroom	3 hours /week= 30 hours/semester	100%
2	Blended	None	0%
3	E-learning	None	0%
4	Distance learning	None	0%
5	Other (Laboratory)	None	0%

7. Contact Hours (based on academic semester)

No	Activity	Contact Hours
1	Lecture	30
2	Laboratory/Studio	N/A
3	Tutorial	N/A
4	Others (specify)	NA
	Total	30 Hours

B. Course Objectives and Learning Outcomes

1. Course Description

This course describes the development, function and regulation of the cells of the immune system in relation to immunological disorders in the human body, and study the methods followed in the clinical laboratories to diagnose these diseases.

2. Course Main Objective

To study the normal and abnormal process associated with immunological disorders and learn various diagnostics approaches applied to immunological clinical laboratories.

3. Course Learning Outcomes

	Aligned PLOs		
1	Knowledge and Understanding		
1.1	Identify the clinical basis of the immune response in normal and abnormal conditions, and the modulation in the immune system components.	K 1	
1.2	Describe the significance of clinical immunological markers and the methods employed in the clinical laboratory.	К2	
2	2 Skills:		
2.1	Assess proper immunological procedures in accurate and precise testing and data analysis.	S2	
3	Values:		
	None		

C. Course Content (Theory)

No	List of Topics	Contact Hours
1	Tolerance of the immune system and Hygiene hypothesis (Presentation)	3
2	Immunizations (Lecture Notes: Immunology/Chapter 12; Pages 353-377)	3
3	Allergy and Hypersensitivity (Lecture Notes: Immunology/Chapter 14; Pages 405-434)	3
4	Tumor Immunology (Lecture Notes: Immunology/Chapter 16; Pages 458-498)	3
5	Immunodeficiency (Lecture Notes: Immunology/Chapter 13; Pages 378-404)	6
6	Autoimmune disorders	6
7	Transplantation (Lecture Notes: Immunology/Chapter9; Pages 458-498)	3
8	Immunotherapy (Presentation)	3
	30	

D. Teaching and Assessment

1. Alignment of Course Learning	Outcomes with	Teaching Strategies	and Assessment Methods
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Code	Course Learning Outcomes	Teaching Strategies	Assessment Methods	
1.0	Knowledge and Understanding			
1.1	Identify the clinical basis of the immune response in normal and abnormal conditions, and the modulation in the immune system components.	• Lectures.	Written ExamAssignment	
1.2	Describe the significance of clinical immunological markers and the methods employed in the clinical laboratory.	• Lectures.	• Written Exam	
2.0	Skills			
2.1	Evaluate proper immunological procedures in accurate and precise testing and data analysis.	LectureProblem-Based Learning	• Written Exam	
3.0	3.0 Values			
	None			

2. Assessment Tasks for Students

#	Assessment task*	Week Due	Percentage of Total Assessment Score
1	Mid-Term Exam	5 th week	30%
2	Activity	8 th week	10%
3	Final Exam12th/13th week		60%
	Total		100%

*Assessment task (i.e., written test, oral test, oral presentation, group project, essay, etc.)

E. Student Academic Counseling and Support

Arrangements for availability of faculty and teaching staff for individual student consultations and academic advice:

- Course instructors are available for individual consultation in their free time. They are usually full-time permanent members present on-campus from 8:00 am to 2:30 pm on all working days. Appointments can be made in person with the instructor through email etc. Days and time availability of each instructor are posted on their doors. Course instructors provide a range of academic and course management advice including course planning and its progression.
- Each student at the department of Clinical Laboratory Sciences has an academic adviser who is available for individual consultation and guidance. Appointments can be made in person with the instructor through email etc. Days and time availability of each adviser are posted on their doors. The academic adviser can provide support with time management, exam preparation, clarification of subject requirements, feedback on performance and dealing with personal issues as well.

F. Learning Resources and Facilities 1.Learning Resources

Required Textbooks	 Todd, I., Spickett, G., & Fairclough, L. (2015). Immunology (7th ed.). Wiley-Blackwell Publishers. ISBN: 978-1-118-45164-9 	
Essential References Materials	• None	
Electronic Materials	• Saudi Digital Library, PubMed, Google Scholar	
Other Learning Materials	 Journals and Articles. 	

2. Facilities Required

Item	Resources
Accommodation (Classrooms, laboratories, demonstration rooms/labs, etc.)	Classrooms
Technology Resources (AV, data show, Smart Board, software, etc.)	Data show, Blackboard and A/V
Other Resources (Specify, e.g. if specific laboratory equipment is required, list requirements or attach a list)	None

G. Course Quality Evaluation

Evaluation Areas/Issues	Evaluators	Evaluation Methods
Student's feedback on effectiveness of teaching.	• Students	• Indirect: Questionnaire Survey at the end of each semester.
Quality of learning resources (laboratory and library) related to each course.	StudentsStaff members	• Indirect: Questionnaire Survey at the end of each semester related to learning resources.
Evaluation of teaching	• Peer evaluators	• Indirect: Peer evaluation
Evaluation of exam quality and assessment.	Exam committeeStudents	 Direct: Exam paper/ exam blueprint review Indirect: Questionnaire Survey at the end of each semester.
Achievement of course learning outcomes	 Course Coordinators Development and accreditation committee 	• Direct: Student's Performance assessed through item analysis and rubrics.

Evaluation areas (e.g., Effectiveness of teaching and assessment, Extent of achievement of course learning outcomes, Quality of learning resources, etc.)

Evaluators (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify) Assessment Methods (Direct, Indirect)

H. Specification Approval Data

Council / Committee	Department Council
Reference No.	Meeting Number 11
Date	19/05/2022





Course Specifications

Course Title:	Diagnostic Parasitology	
Course Code:	373321-3	
Program:Bachelor in Clinical Laboratory Sciences; Letter (Program Code: 373000)		
Department:	Clinical Laboratory Sciences Department	
College:	College of Applied Medical Sciences	
Institution:	Taif University	







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1.Learning Resources	7
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A.	A. Course Identification				
1.	1. Credit hours: 3 Hours (2 T + 1 P)				
2. a. b.					
3.	3. Level/year at which this course is offered: 7 th Level/ Third Year				
4. Pre-requisites for this course (if any): Basic of Medical Microbiology/ 373228-3					
5. Co-requisites for this course (if any): None					

6. Mode of Instruction (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1	Traditional classroom	3 hours /week= 30 hours/semester	50%
2	Blended	None	0%
3	E-learning	None	0%
4	Distance learning	None	0%
5	Other (Laboratory)	3 hours /week= 30 hours/semester	50%

7. Contact Hours (based on academic semester)

No	Activity	Contact Hours
1	Lecture	30
2	Laboratory/Studio	30
3	Tutorial	N/A
4	Others (specify)	NA
	Total	60 Hours

B. Course Objectives and Learning Outcomes

1. Course Description

The course is intended to give students concepts about types of parasites that infect humans, their pathogenicity together with a brief clinical description and determining suitable clinical specimens for laboratory diagnosis using microscopy, immunoassays as well as molecular diagnosis.

2. Course Main Objective

By the end of this course, students will gain adequate knowledge regarding human parasites in terms of distinct classes, pathology, pathogenicity, transmission and laboratory diagnosis and will be able to select proper diagnostic tests and interpret the results. They will also be able to do different parasitological diagnostic techniques both independently and in groups.

3. Course Learning Outcomes

	CLOs	Aligned PLOs	
1	Knowledge and Understanding		
1.1	Describe pathogenesis of underlying infections with different parasites of medical importance to human in terms of their classification, morphology and pathology.	K1	
1.2	1.2 Identify the most appropriate clinical specimen(s) and laboratory test(s) used for diagnosis of different parasitic infections.		
2	Skills:		
2.1	Perform different parasitological laboratory tests in safe and effective way.	S1	
2.2	Interpret laboratory test finding(s) and correlate it (them) with the patient's clinical picture(s).	S2	
3	Values:		
3.1	Demonstrate self-learning ability during research in library, and internet as a source of knowledge to prepare assignments and perform tasks.	V1	
3.2	Exhibit responsibility by creating awareness among community in areas related to the course.	V2	

C. Course Content

No	List of Theory Topics	Contact Hours
1	Introduction to Diagnostic Parasitology (Presentation)	3
	• Terms	
	• Type of parasites	
	• Type of hosts	
	• pathology, pathogenesis	
2	Protozoa: (Presentation)	3
	General Characters	
	Classification	
	• Ameobiasis	
	• Giardiasis	
	Coccidian parasites	
	• Trichomoniasis .	
3	Extra-intestinal protozoa: (Presentation)	3
	• Leishmania	
	• Toxoplasma	
4	Plasmodium (Book Chapter 4; Page 79)	3
	• Epidemiology	
	• Life cycle	

	Clinical picture	
	• Pathogenesis	
	Laboratory diagnosis	
5	Trematodes: (Book Chapter 6; Pages 166 and 181)	3
	General Characteristics of flat worms	
	Schistosomes	
6	Trematodes: (Presentation)	3
	• liver flukes	
	Intestinal flukes	
7	Cestodes: (Presentation)	3
	General Characteristics	
	• FishTapeworm,	
	• Taeniasis	
	Cysticercosis	
8	Cestodes: (Book Chapter 7; Page 207)	3
	Hydatid cyst	
	Hymenolepsiasis	
9	Nematodes: (Book Chapter 8; Page 239)	3
	General Characters	
	Classification	
	Ascaris Lumbricoides	
	• Hookworm	
	• Whipworm	
10	Nematodes: (Presentation)	3
	• pinworm infections	
	• Threadworm	
	Lymphatic filariasis	
	Total	30

No	List of Practical Topics	Contact Hours	
1	Laboratory safety		
	• Samples	3	
_	• Sampling and microscopy.		
	(Lab Manual) Demonstration slides of:		
	Ameobiasis		
	Giardiasis		
2	Coccidian parasites	3	
	 Trichomoniasis. 		
	(Lab Manual)		
	Demonstration slides of :		
	• Leishmania	2	
3	• Toxoplasma.	3	
	(Lab Manual)		
	Demonstration slides of :		
4	• Plasmodium	3	
	(Lab Manual)		
	Stool concentration		
_	Staining techniques		
5	 Demonstration slides(Schistosomes) 	3	
	(Lab Manual)		
	Trematodes:		
6	• liver flukes	3	
	Intestinal flukes		
	(Lab Manual)		
	Demonstration slides of:		
	Fish Tapeworm Transie	2	
7	Taeniasis Cautionnesis	3	
	• Cysticercosis		
┝──┤	(Lab Manual) Demonstration slides of:		
	Hydatid cyst		
8	 Hydalid Cyst Hymenolepsiasis 	3	
	(Lab Manual)		
┝──┤	Demonstration slides of:		
	Ascaris Lumbricoides		
9	Hookworm	3	
,	• Whipworm	-	
	(Lab Manual)		
	Demonstration slides of:	3	
	Pinworm infections		
10	• Threadworm		
10	Lymphaticfilariasis		
	(Lab Manual)		
	Total 30		
	10(a)	30	

D. Teaching and Assessment

1. Alignment of Course Learning Outcomes with Teaching Strategies and Assessment Methods

Code	Course Learning Outcomes	Teaching Strategies	Assessment Methods
1.0	Knowledge and Understanding		
1.1	Describe pathogenesis of underlying infections with different parasites of medical importance to human in terms of their classification, morphology and pathology.	• Lecture	• Written Exam
1.2	Identify the most appropriate clinical specimen(s) and laboratory test(s)used for diagnosis of different parasitic infections.	LecturePractical Session	Written ExamPractical ExamLab Report
2.0	Skills		
2.1	Perform different parasitological laboratory tests in safe and effective way.	Practical Session	 Practical Exam Lab Report
2.2	Interpret laboratory test finding(s) and correlate it (them) with the patient's clinical picture(s).	 Lectures Practical Session Problem based learning. 	Written ExamsOSPE
3.0	Values		
3.1	Demonstrate self-learning ability during research in library, and internet as a source of knowledge to prepare assignments and perform tasks.	Group Discussion	• Activity
3.2	Exhibit responsibility by creating awareness among community in areas related to the course.		• Activity

#	Assessment task*	Week Due	Percentage of Total Assessment Score
1	Mid-Term Exam	5 th week	15%
2	Activity	Throughout	5%
3	Practical Report	Throughout	10%
4	Final Practical Exam	11 th week	20%
5	Final Exam	12 th /13 th week	50%
	Total		100%

2. Assessment Tasks for Students

*Assessment task (i.e., written test, oral test, oral presentation, group project, essay, etc.)

E. Student Academic Counseling and Support

Arrangements for availability of faculty and teaching staff for individual student consultations and academic advice:

- Course instructors are available for individual consultation in their free time. They are usually full-time permanent members present on-campus from 8:00 am to 2:30 pm on all working days. Appointments can be made in person with the instructor through email etc. Days and time availability of each instructor are posted on their doors. Course instructors provide a range of academic and course management advice including course planning and its progression.
- Each student at the department of Clinical Laboratory Sciences has an academic adviser who is available for individual consultation and guidance. Appointments can be made in person with the instructor through email etc. Days and time availability of each adviser are posted on their doors. The academic adviser can provide support with time management, exam preparation, clarification of subject requirements, feedback on performance and dealing with personal issues as well.

F. Learning Resources and Facilities

1.Learning Resources

Required Textbooks	 David T. John, William A. Petri. Markell and Voge's Medical Parasitology (2006), 9th Edition, Elsevier Inc., Saunders Imprint. ISBN-13: 978-0-721-64793-7; ISBN-10: 0-7216-4793-6.
Essential References Materials	 Julia Walochnik and Michael Duchene, Molecular Parasitology (2016), 1st Edition, Springer Publishers, ISBN: 978-3-7091-1416-2. Elizabeth A. Zeibig. Clinical Parasitology: A Practical Approach (2013), 2nd Edition. Elsevier Saunders, ISBN: 978-1-4160-6044-4
Electronic Materials	None
Other Learning Materials	Websites, Search engines (Saudi Digital Library, PubMed, Google Scholar)

2. Facilities Required

Item	Resources
Accommodation (Classrooms, laboratories, demonstration rooms/labs, etc.)	Classrooms and Laboratories
Technology Resources (AV, data show, Smart Board, software, etc.)	Data show, Blackboard and A/V
Other Resources (Specify, e.g. if specific laboratory equipment is required, list requirements or attach a list)	N/A

G. Course Quality Evaluation

Evaluation Areas/Issues	Evaluators	Evaluation Methods
Student's feedback on effectiveness of teaching.	• Students	• Indirect: Questionnaire Survey at the end of each semester.
Quality of learning resources (laboratory and library) related to each course.	StudentsStaff members	• Indirect: Questionnaire Survey at the end of each semester related to learning resources.
Evaluation of teaching	• Peer evaluators	• Indirect: Peer evaluation
Evaluation of exam quality and assessment.	Exam committeeStudents	 Direct: Exam paper/ exam blueprint review Indirect: Questionnaire Survey at the end of each semester.
Achievement of course learning outcomes	 Course Coordinators Development and accreditation committee 	• Direct: Student's Performance assessed through item analysis and rubrics.

Evaluation areas (e.g., Effectiveness of teaching and assessment, Extent of achievement of course learning outcomes, Quality of learning resources, etc.)

Evaluators (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify) Assessment Methods (Direct, Indirect)

H. Specification Approval Data

Council / Committee	Department Council
Reference No.	Meeting Number 11
Date	19/05/2022





Course Specifications

Course Title:	Hematology 1
Course Code:	373313-3
Program:	Bachelor in Clinical Laboratory Sciences; Level-6 (Program Code: 373000)
Department:	Clinical Laboratory Sciences Department
College:	College of Applied Medical Sciences
Institution:	Taif University







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1.Learning Resources	7
2. Facilities Required	7
G. Course Quality Evaluation7	
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A. Course Identification

1.	1. Credit hours: 3 Hours (2 T + 1 P)		
2. 0	Course type		
a.	University College Department Others		
b.	Required Elective		
3.]	Level/year at which this course is offered: 7th Level/ Third Year		
4.]	4. Pre-requisites for this course (if any): Human Physiology/ 373220-2		
5.	5. Co-requisites for this course (if any): None		

6. Mode of Instruction (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1	Traditional classroom	3 hours/week= 30 hours/semester	50%
2	Blended	None	0%
3	E-learning	None	0%
4	Distance learning	None	0%
5	Other (Laboratory)	3 hours /week= 30 hours/semester	50%

7. Contact Hours (based on academic semester)

No	Activity	Contact Hours
1	Lecture	30
2	Laboratory/Studio	30
3	Tutorial	N/A
4	Others (specify)	NA
	Total	60 Hours

B. Course Objectives and Learning Outcomes

1. Course Description

This course is intended to provide the students with the basic concepts of the generation, structure, function and metabolism of red blood cells. Moreover, it is a combined lecture and laboratory course covering methods for analysis of blood cells, the classification and laboratory investigation of anemias and other common disorders involving red blood cells.

2. Course Main Objective

The main purpose of this course is directed towards the learning and understanding of pathological disorders of red blood cells, including anemias and other common disorders involving red blood cells.

3. Course Learning Outcomes

CLOs		Aligned PLOs
1	Knowledge and Understanding	
1.1	Explain red blood cell disorders by relating the etiology, pathophysiology and laboratory diagnosis of various hematological disorders.	K1
1.2	Recognize principles applied for hematology laboratory settings including specimen collection and suitability, instrumentation, testing and reporting	К2
2	Skills	
2.1	Perform basic red blood cells hematological techniques used in a diagnostic laboratory in a safe and effective manner	S1
2.2	Analyze clinical cases and interpret the red blood cells hematological results to determine a final diagnosis.	S2
2.3	Apply effective communication of scientific and hematological concepts clearly, concisely and logically.	S4
3	Values	
3.1	Display professional and responsible attitude while performing tasks related to the course.	V1

C. Course Content (Theory)

No	List of Topics	Contact Hours
1	Introduction to hematology (Presentation)	3
2	Hematopoiesis (Presentation)	3
3	Erythrocytes structure and metabolism (Presentation)	3
4	Hemoglobin Function and structure (Presentation)	3
5	Introduction to Anemia (Presentation)	3
6	Anemias of Disordered Iron Metabolism and Heme Synthesis (Presentation)	3
7	Hemoglobinopathies (Presentation)	3
8	Megaloblastic and Nonmegaloblastic Macrocytic Anemias (Presentation)	3
9	Hemolytic Anemia: Membrane Defects and Enzyme Deficiencies (Presentation)	3
10	Hemolytic Anemia: Immune Anemias and Nonimmune Defects (Presentation)	3
	Total	30

C. Course Content (Practical)

No	List of Topics	Contact Hours
1	Introduction & Universal precautions (Lab Manual)	3
2	Hemoglobin Estimation Sahali method + HICN method. (Lab Manual)	6
3	Hemocytometry & Total erythrocyte count (Lab Manual)	3
4	Determination of Haematocrit (paced cell volume (Lab Manual)	3
5	Absolute values (red cell indicies) MCV, MCH, MCHC (Lab Manual) (Lab Manual)	3
6	Preparation, staining and examination of blood film 1 (Lab Manual)	3
7	Preparation, staining and examination of blood film 2 (Lab Manual)	3
8	Reticulocyte count (Lab Manual)	3
9	Introduction & Universal precautions (Lab Manual)	3
10	Normal & abnormal morphology of red blood cells (Lab Manual)	3
Total		

D. Teaching and Assessment

1. Alignment of Course Learning Outcomes with Teaching Strategies and Assessment Methods

Code	Course Learning Outcomes	Teaching Strategies	Assessment Methods
1.0	Knowledge and Understanding		
1.1	Explain red blood cell disorders by relating the etiology, pathophysiology and laboratory diagnosis of various hematological disorders.	• Lecture	• Written Exam
1.2	Recognize principles applied for hematology laboratory settings including specimen collection and suitability, instrumentation, testing and reporting.	LecturePractical Session	Written ExamPractical ExamLab Report
2.0	Skills		
2.1	Perform basic red blood cells hematological techniques used in a diagnostic laboratory in a safe and effective manner.	Practical Session	Practical ExamLab Report
2.2	Analyze clinical cases and interpret the red blood cells hematological results to determine a final diagnosis.	 Lecture Practical Session Problem- Based Learning 	Written ExamOSPE

Code	Course Learning Outcomes	Teaching Strategies	Assessment Methods
2.3	Apply effective communication of scientific and hematological concepts clearly, concisely and logically.	Group Discussion	Written ExamScientific Activity
3.0	Values		
3.1	Display professional and responsible attitude while performing tasks related to the course.	Practical Session	• Practical Exam

2. Assessment Tasks for Students

#	Assessment task*	Week Due	Percentage of Total Assessment Score
1	Mid-Term Exam	5 th week	15%
2	Activity	Throughout	5%
3	Practical report	Throughout	10%
4	Final Practical Exam	11 th week	20%
5	Final Exam	12 th /13 th week	50%
	Total		100%

*Assessment task (i.e., written test, oral test, oral presentation, group project, essay, etc.)

E. Student Academic Counseling and Support

Arrangements for availability of faculty and teaching staff for individual student consultations and academic advice:

- Course instructors are available for individual consultation in their free time. They are usually full-time permanent members present on-campus from 8:00 am to 2:30 pm on all working days. Appointments can be made in person with the instructor through email etc. Days and time availability of each instructor are posted on their doors. Course instructors provide a range of academic and course management advice including course planning and its progression.
- Each student at the department of Clinical Laboratory Sciences has an academic adviser who is available for individual consultation and guidance. Appointments can be made in person with the instructor through email etc. Days and time availability of each adviser are posted on their doors. The academic adviser can provide support with time management, exam preparation, clarification of subject requirements, feedback on performance and dealing with personal issues as well.

F. Learning Resources and Facilities 1.Learning Resources

	• Shirlyn McKenzie, Kristin Landis-Piwowar, Linne Williams
Required Textbooks	 Shiriyi McKenzle, Kristin Landis-Piwowar, Linne Williams (2019). <i>Clinical Laboratory Hematology</i>, 4th Edition, Pearson Publishers. ISBN-13: 978-0134709390; ISBN-10: 013470939X Jacquelline H. Carr (2021). <i>Clinical haematology Atlas</i>, 6th Edition, Elsevier Publishers. ISBN-13: 978-0323711920; ISBN-10: 0323711928 Victor Hoffbrand and David Steensma (2019). <i>Hoffbrand's Essentials: Haematology</i>, 8th Edition. John Wiley and Sons Ltd. ISBN-13: 978-1119495901; ISBN-10: 1119495903
Essential References Materials	• None
Electronic Materials	• None
Other Learning Materials	Saudi Digital Library

2. Facilities Required

Item	Resources
Accommodation (Classrooms, laboratories, demonstration rooms/labs, etc.)	Classrooms and Laboratories
Technology Resources (AV, data show, Smart Board, software, etc.)	Data show, Blackboard and A/V
Other Resources (Specify, e.g. if specific laboratory equipment is required, list requirements or attach a list)	Complete blood count machineLight MicroscopesHb Electrophoresis

G. Course Quality Evaluation

Evaluation Areas/Issues	Evaluators	Evaluation Methods
Student's feedback on effectiveness of teaching.	• Students	• Indirect: Questionnaire Survey at the end of each semester.
Quality of learning resources (laboratory and library) related to each course.	StudentsStaff members	• Indirect: Questionnaire Survey at the end of each semester related to learning resources.
Evaluation of teaching	• Peer evaluators	• Indirect: Peer evaluation

Evaluation Areas/Issues	Evaluators	Evaluation Methods
Evaluation of exam quality and assessment.	Exam committeeStudents	 Direct: Exam paper/ exam blueprint review Indirect: Questionnaire Survey at the end of each semester.
Achievement of course learning outcomes	 Course Coordinators Development and accreditation committee 	• Direct: Student's Performance assessed through item analysis and rubrics.

Evaluation areas (e.g., Effectiveness of teaching and assessment, Extent of achievement of course learning outcomes, Quality of learning resources, etc.)

Evaluators (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify) Assessment Methods (Direct, Indirect)

H. Specification Approval Data

Council / Committee	Department Council
Reference No.	Meeting Number 11
Date	19/05/2022





Course Specifications

Course Title:	Toxicology
Course Code:	373324-2
Program:	Bachelor in Clinical Laboratory Sciences; Level-6 (Program Code: 373000)
Department:	Clinical Laboratory Sciences Department
College:	College of Applied Medical Sciences
Institution:	Taif University







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A. Course Identification

1.	1. Credit hours: 2 Hours (1 T + 1 P)		
2. (Course type		
a.	University College Department Others		
b.	Required Elective		
3.	Level/year at which this course is offered: 7 th Level/ Third Year		
4.	Pre-requisites for this course (if any): None		
5.	5. Co-requisites for this course (if any): None		

6. Mode of Instruction (mark all that apply)

No	Mode of Instruction	Contact Hours	Percentage
1	Traditional classroom	2 hours /week = 20 hours/semester	40%
2	Blended	None	0%
3	E-learning	None	0%
4	Distance learning	None	0%
5	Other (Laboratory)	3 hours /week= 30 hours/semester	60%

7. Contact Hours (based on academic semester)

No	Activity	Contact Hours
1	Lecture	20
2	Laboratory/Studio	30
3	Tutorial	N/A
4	Others (specify)	NA
	Total	50 Hours

B. Course Objectives and Learning Outcomes

1. Course Description

Toxicology covers the studying of poisonous chemicals, drugs, carcinogens, and other exogenous compounds. The adverse effects of these chemicals in each body system, as well as their metabolism and detoxification from the body will be covered in this course.

The course will also cover dose/effect relationships and route of exposure in addition to public health implications and types of toxicology (Analytic/forensic, clinical, environmental and food toxicology).

2. Course Main Objective

The student will know, interpret and explain concepts of toxicology and how to solve complex toxicological emergency situations using critical thinking skills. The student will also be able to design and formulate experiments to test or challenge hypotheses presented in the toxicology laboratory.

3. Course Learning Outcomes

	CLOs	Aligned- PLOs
1	Knowledge and Understanding	
1.1	Describe the effect of the chronic and acute exposures to toxic chemicals on different body systems, and the toxicological mechanisms with special emphasis on their dose response, metabolism and detoxification.	K1
1.2	Identify principles of applied clinical toxicological laboratory including instrumentation, methods, and results interpretation for targeted organs and indicators.	
2	Skills:	
2.1	Perform sample preparation for toxicological analysis, analytical practice for skills in methodology and post-analytical interpretive conclusive remarks.	S1
2.2	Interpret laboratory data and correlate it with clinical manifestation of toxic chemical type.	S2
3	Values:	
3.1	Display knowledge on toxicological adverse effect of drug abuse through increasing community awareness.	V2

C. Course Content (Theory and Practical)

No	List of Topics (THEORY)	Contact Hours
1	Principles of toxicology (Book Ch.2, P. 11) Definitions, areas, abbreviations of toxicology Risk, safety and tolerance Types of Chemical Interactions Adverse effects types Drug development and toxicity Dose response relationship Therapeutic index and toxicity rating Why do we need toxicity testing? 	2
2	 Mechanisms of toxicology (Book Ch.3, P. 45) Allergic response types Receptor, ion channel and enzyme-mediated toxicity Biochemical pathways and organ-directed toxicity Mutagenesis, carcinogenesis, and teratogenicity Snake Bites in Saudi Arabia and its antidotes 	2

3	 Toxicokinetics (Book Ch.7, P. 305) Absorption and bioavailability Distribution Biotransformation Excretion and Clearance Importance of Toxicokinetics and its applications 	2
4	 Agonists- antagonists & Dote-antidote (Presentation) Agonists & Antagonists Competitive & non-competitive antagonists Antidotes Poisoning in Saudi Arabia Antidotes available in emergency department & pharmacy 	2
5	 Organ Toxicity: Liver toxicity (Book Ch.13, P. 557) Liver structure & functions Types of liver injury and its laboratory indications Mechanisms of hepatoxicity Types of hepatoxicants & its metabolic activation 	2
6	 Organ Toxicity: Kidney toxicity (Book Ch.14, P. 583) Kidney structure & functions Types of kidney injury and its laboratory indications Mechanisms of nephrotoxicity Types of nephrotoxicants 	2
7	 Organ Toxicity: Respiratory and nervous toxicity (Presentation) Anatomy & functions of respiratory system Respiratory system injury and its laboratory indications Occupational & environmental lung diseases/cancer Toxicants that induce respiratory system injury & remodeling Anatomy & functions of nervous system Toxicants effects on nervous system (Demyelination, axonopathy, neuronopathy, & impaired Synaptic Function) In vivo & in vitro tests for neurotoxicity 	2
8	 Environmental toxicology (Book Ch.29, P. 1117) Heavy metals Pesticides & insecticides Environmental hazardous waste & its effect on health Effects of environmental hazards on respiratory tract, skin, eye, nervous system, liver, and kidneys 	2

9	 Food and domestic toxicology (Book Ch.30, P. 1191) Safety Requirements for Dietary Supplements Adverse Reactions to Food & laboratory indications Microbiological Agents in food Toxins in Fish & seafood Toxins in processed food (contamination in storage/processing) Household hazardous materials (cleaning products, pesticides, painting supplies, batteries, cosmetics & drugs) Emergency management 	
10	 Forensic toxicology (Presentation) Criminal poisoning Toxicologic investigation of a poison death Forensic drug abuse testing Illegal athlete drugs Interpretation of Analytic Results 	2
	Total theoretical hours	
No	List of Topics (PRACTICAL)	
	 Introduction: Methods in toxicology I (Lab Manual) Clinical and preclinical toxicological investigations Cell culture techniques 	
11	 Cell culture instruments Inverted microscope, fluorescent microscope, laminar flow, CO₂ incubator, ect) Cells separation from body fluids Cell toxicity techniques 	3

13	 Classification of samples and toxicological tests (Lab Manual) Containers for specimens collection Tier I toxicology testing Stat quantitative serum & urine toxicology assays Tier II toxicology testing Tests for the diagnosis of any poisoning/toxicity (toxicological, hematological, microbiological, serological, clinical, cytological, histological, genetic tests and body fluid analysis) 	3
14	 Emergency poisoning symptoms (Lab Manual) General poisoning symptoms Specific poisoning symptoms Vomiting color meaning and causes 	3
15	 Common drugs abuse tests (Lab Manual) Classification of common drugs of abuse Specimens for drug abuse detection Turnaround time for each drug Tests, kits, and instruments to detect drugs of abuse 	3
16	 Analytical toxicology tests I (Lab Manual) Electrolytes types Anion gap and its laboratory indications Electrolytes defect types Osmolar gap and its laboratory indications 	3
17	 Analytical toxicology tests II (Lab Manual) Toxicological screening tests Glucose level, liver function tests, urine analysis, Blood Creatine Kinase, Blood urea nitrogen (BUN) and creatinine Chemical Spot Tests Electrocardiogram Gas chromatographic screening for drugs , Thin layer chromatography, HPLC Direct (colorimetric tests) in serum or urine sample Toxicity for specific drug 	3

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18	 Therapeutic Drug Monitoring (Lab Manual) TDM definition & concept When TDM is required? Drugs that do not need TDM Commonly monitored drugs TDM for Aminoglycosides Guidelines for specimen collection and timing for TDM Interpretive criteria Factors that affect TDM results Reasons for pharmacokinetic variability Analytical methods for TDM Clinical significance of TDM 	3
19	 Metal poisoning detection (Lab Manual) Metal Poisoning symptoms & specimens Methods for detection of metal poisoning in human specimens for different metals. Instruments for metal poisoning detection Antidote for metal poisoning 	3
20	 Genetic toxicity tests (Lab Manual) Genotoxicity types Genotoxic agents Genotoxicity tests Instruments to detect genotoxicity (RT-PCR, next-generation sequencing, and confocal microscope) Molecular Testing: small DNA mutations, single nucleotide polymorphism, DNA Fingerprinting, Variable Number Tandem Repeats (VNTR) Analysis Cytogenetic tests: comet test, micronucleus assays, karyotyping Tests for genetic disorders (Homogeneous disorders, heterogeneous disorders, chromosomal numerical defect syndromes, and chromosomal aberrations or rearrangements 	3
	Total practical hours	30

D. Teaching and Assessment

1. Alignment of Course Learning Outcomes with Teaching Strategies and Assessment Methods

Code	Course Learning Outcomes	Teaching Strategies	Assessment Methods
1.0	Knowledge and Understanding		
1.1	Describe the effect of the chronic and acute exposures to toxic chemicals on different body systems, and the toxicological mechanisms with special emphasis on their dose response, metabolism and detoxification.	• Lecture	• Written Exam
1.2	Identify principles of applied clinical toxicological laboratory including instrumentation, methods, and results interpretation for targeted organs and indicators.	LecturePractical Session	Written ExamPractical ExamLab Report
2.0	Skills	/	
2.1	Perform sample preparation for toxicological analysis, analytical practice for skills in methodology and post-analytical interpretive conclusive remarks.	Practical session	 Practical Exam Lab Report
2.2	Interpret laboratory data and correlate it with clinical manifestation of toxic chemical type.	 Lecture Practical Session Problem-Based Learning 	• Written Exam
4.0	Values		
4.1	Display responsible attitude through increasing community awareness on toxicological adverse effect of drug abuse.	• Service Learning	• Activity

2. Assessment Tasks for Students

#	Assessment task*	Week Due	Percentage of Total Assessment Score
1	Mid-Term Exam	4 th Week	15%
2	Practical Report	6 th Week	10%
3	Activity	8 th Week	5%
4	Final Practical Exam	11 th Week	20%
5	Final Exam	12 th Week	50 %
	Total		100%

*Assessment task (i.e., written test, oral test, oral presentation, group project, essay, etc.)

E. Student Academic Counseling and Support

Arrangements for availability of faculty and teaching staff for individual student consultations and academic advice:

- Course instructors are available for individual consultation in their free time. They are usually full-time permanent members present on-campus from 8:00 am to 2:30 pm on all working days. Appointments can be made in person with the instructor through email etc. Days and time availability of each instructor are posted on their doors. Course instructors provide a range of academic and course management advice including course planning and its progression.
- Each student at the department of Clinical Laboratory Sciences has an academic adviser who is available for individual consultation and guidance. Appointments can be made in person with the instructor through email etc. Days and time availability of each adviser are posted on their doors. The academic adviser can provide support with time management, exam preparation, clarification of subject requirements, feedback on performance and dealing with personal issues as well.

1.Learning Kesourt	
Required TextbooksCasarett & Doull's Toxicology: The Basic Science of Poin Edition, 2019 McGraw-Hill, Medical Publishing Division 13: 978-125986374	
	• Barbarajean Magnani, Michael G. Bissell, Tai C. Kwong, Alan H.B. Wu, CLINICAL TOXICOLOGY TESTING: A GUIDE FOR LABORATORY PROFESSIONALS - PUB220. ISBN: 978-0- 9837068-1-6
	• Frank A. Barile. Barile's Clinical Toxicology Principles and Mechanisms. 3rd Edition 2019, CRC Press (Taylor and Francis Group) ISBN: 9781498765305.
Essential References Materials	• CASARETT AND DOULL'S Toxicology. 7th Edition. Curtis D. Klaassen, 2008, McGraw-Hill, Medical Publishing Division.
Electronic Materials	http://ban1080.org.au/wp- content/uploads/2019/11/casarett_and_doull_s_toxicology- the_basic_science_of_poisons_7th_edition_2008.pdf
Other Learning Materials	• None

F. Learning Resources and Facilities 1. Learning Resources

2. Facilities Required

Item	Resources
Accommodation (Classrooms, laboratories, demonstration rooms/labs, etc.)	Classrooms and Laboratories
Technology Resources (AV, data show, Smart Board, software, etc.)	Data show, Blackboard and A/V
Other Resources (Specify, e.g. if specific laboratory equipment is required, list requirements or attach a list)	Microtiter reader for absorbance, centrifuge, incubator, fluorescence microscope, real-time thermocycler, electrophoresis system, and pipettes.

G. Course Quality Evaluation

Evaluation Areas/Issues	Evaluators	Evaluation Methods
Student's feedback on effectiveness of teaching.	• Students	• Indirect: Questionnaire Survey at the end of each semester.
Quality of learning resources (laboratory and library) related to each course.	StudentsStaff members	• Indirect: Questionnaire Survey at the end of each semester related to learning resources.
Evaluation of teaching	• Peer evaluators	• Indirect: Peer evaluation
Evaluation of exam quality and assessment.	Exam committeeStudents	 Direct: Exam paper/ exam blueprint review Indirect: Questionnaire Survey at the end of each semester.
Achievement of course learning outcomes	 Course Coordinators Development and accreditation committee 	• Direct: Student's Performance assessed through item analysis and rubrics.

Evaluation areas (e.g., Effectiveness of teaching and assessment, Extent of achievement of course learning outcomes, Quality of learning resources, etc.)

Evaluators (Students, Faculty, Program Leaders, Peer Reviewer, Others (specify) Assessment Methods (Direct, Indirect)

H. Specification Approval Data

Council / Committee	Department Council
Reference No.	Meeting Number 11
Date	19/05/2022

